Capan-1

300143

GENERAL INFORMATION

Description

The Capan-1 cell line is derived from a human pancreatic adenocarcinoma and was established from the ascitic fluid of a 40-year-old Caucasian male. It was first characterized in 1975 and is particularly noted for its ductal epithelial morphology, which closely resembles that of primary pancreatic tumors. Capan-1 cells are extensively used in research aimed at understanding pancreatic cancer biology, including studies on tumor progression, metastasis, and treatment resistance. This cell line is well-regarded for its ability to produce mucin, a characteristic feature of many pancreatic adenocarcinomas, thus serving as a model for mucinous pancreatic cancer.
Genetically, Capan-1 harbors mutations in the KRAS gene, which are typical of pancreatic cancer, as well as alterations in other cancer-related genes such as TP53 and SMAD4. These mutations make the Capan-1 cell line a valuable tool for studying the molecular mechanisms underlying pancreatic cancer and for the preclinical evaluation of new therapeutic agents targeting these pathways. Furthermore, Capan-1 cells are used to study the biology of pancreatic cancer stem cells, offering insights into the behaviors that drive cancer recurrence and resistance to conventional therapies.

Tissue	Pancreas
Disease	Ductal adenocarcinoma
Metastatic site	Liver

Synonyms

- CaPan-1
- CAPAN-1
- Capan 1
- CAPAN 1
- Capan1
- CAPAN1

CLASSIFICATION

Organism	Human
Sex	Male
Growth properties	Adherent
Biosafety level	1

CHARACTERISTICS

Morphology	Epithelial-like
Tumourigenic	Form adenocarcinoma consistent with pancreatic duct carcinoma

Karyotype

(P7) hypotriploid with abnormalities including dicentrics, breaks, acrocentric fragments, large submetacentric and subtelocentric chromosomes plus minute marker

Doubling time	60 to 80 hours

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IDENTIFIERS

Citation

Capan-1 (Cytion catalog number 300143)

NCBI TaxID	9606
Cellosaurus	CVCL_0237

EXPRESSION AND SUSCEPTIBILITY

Protein expression

P53 negative

Antigen expression

Blood Type A, Rh+

Isoenzymes

Me-2, 1, PGM3, 1, PGM1, 1-2, ES-D, 1, AK-1, 1, G6PD, B, GLO-1, 1-2, Phenotype Frequency Product: 0.0311

Mutational profile

Capan-1 cells carry a homozygous Kras mutation in codon12: GGT(Gly) >GTT(Val)

Products

Mucin

HANDLING AND CULTURE

Culture medium

RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% FBS

Dissociation reagent	Accutase

Subculturing

Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

Seeding density	2 x 10^4 cells/cm^2 will result in a 90% confluent monolayer in about 7 days
Fluid renewal	Every 3 days
Post-thaw recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 48 hours.

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	40 years

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes