HEK293 □□

300192

HEK293 □□□□ 20 □□ 70 □□□□□□□□□□□□□□□-□□□□□Alex van der Eb□□□□□□□□□□□□□□

Growth properties	Adherent
GMO status	GMO-S1

GMO status details

GMO-S1: This HEK293 embryonic kidney-derived cell line contains adenovirus-5 E1A/E1B sequences due to transformation but does not release infectious virus, enabling high proliferative capacity. The modification is stably present in embryonal kidney cells. This classification applies only within Germany and may differ elsewhere.

Biosafety level	1

CHARACTERISTICS

Morphology	□□□
Tumourigenic	□□
Ploidy status	30% □ HEK293 □□□□ 64 □□□□□□□□□□□□□□4.2%□□□□□□□□□□□□
Doubling time	30 □□

IDENTIFIERS

Citation

HEK293□Cytion □□□ 300192□

NCBI TaxID	9606
Cellosaurus	CVCL_0045

EXPRESSION AND SUSCEPTIBILITY

Receptors expressed

Vitronectin

Protein expression

CEA □□□p53 □□

Virus susceptibility

□□□□ 5 DNA □□□□ □□□ 5 DNA

HANDLING AND CULTURE

Culture medium

EMEM□MEM Eagle□□w□2 mM L-□□□□□w□2.2 g/L NaHCO3□w□EBSS□Cytion □□□□ 820100a□□

Incubation atmosphere	37°C, 5%CO2, □□□□□

Supplements

□□□□□□□ 10% FBS □ 1% NEAA

Dissociation reagent	Accutase

Subculturing

□□□□□□□□□□□□□□□□□□□□□ PBS □□□T25 □□□ 3-5 □□ PBS□T75 □□□ 5-10 □□□□□□ Accutase □□□□□□□T25 □□□ 1-2 □□□T75 □□□ 2.5 □□□□□□□□□□□□ 8-10 □□□□□□□□□□□□□ 10 □□□□□□□□□□□□□□□□□□□□ 300xg □□ 3 □□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□

Seeding density	1×10⁴□□□/cm²□□□4□□□□□□□□

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BIC: SOLADES1HDB

Fluid renewal	□□ 2 □
Post-thaw recovery	□□□□□□□□5×10⁴□□□/□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□24□□□

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

□□□□ PCR □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□
□□

Freeze medium	□□□□□□□□□□□□□□□ FBS□+10% DMSO □ CM-1□Cytion □□□□□□ 800100□□□□□□□□□ □□□□□□□□□□□□□□□□□CM-1□Cytion □□□□□□ 800100□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□

Storage conditions

□□□□□□□□□□□□□□□□□□□□□□□□□□ -150 □ -196 ℃□□-80 °C□□□□□□□□□□□□□□□□□□□□

Shipping conditions

□□□ -78 °C □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□

DONOR

Donor age	□□

LEGAL AND SAFETY

Safety precautions

□□□□□□□□□□□□□□□□□□□