Kasumi-1

300226

GENERAL INFORMATION

Description

The Kasumi-1 cell line was derived from the peripheral blood of a 7-year-old Japanese boy with acute myeloid leukemia (AML), specifically the FAB M2 subtype, during a relapse following bone marrow transplantation. This cell line is a valuable resource for researchers studying hematologic malignancies, especially those involving the t(8;21) chromosomal translocation. This translocation leads to the formation of the AML1-ETO fusion gene, a critical factor in certain subtypes of AML. Kasumi-1 cells thus serve as an essential model for investigating the molecular mechanisms of AML and testing potential therapeutic approaches.
Kasumi-1 cells possess characteristics of both myeloid and macrophage lineages, making them particularly useful for studies on myeloid differentiation. These cells can be induced to differentiate into macrophage-like cells when cultured with phorbol 12-myristate 13-acetate (TPA), providing a robust system for exploring the pathways involved in myeloid lineage commitment and differentiation. This differentiation capacity enhances the utility of Kasumi-1 cells in research focused on both AML biology and broader myeloid cell development processes.

Tissue	Blood
Disease	Acute myeloblastic leukemia

Synonyms

- KASUMI-1
- Kasumi 1
- KASUMI1
- Kasumi1

CLASSIFICATION

Organism	Human
Sex	Male
Growth properties	Suspension
Biosafety level	1

CHARACTERISTICS

Morphology	Round cells showing marked variations in both size and nuclear cytoplasmic ratio.
Cell type	Myeloblast (AML-acute myeloid leukemia)

Karyotype

T(8,21) chromosome translocation

Doubling time	40 to 45 hours

IDENTIFIERS

Citation

Kasumi-1 (Cytion catalog number 300226)

NCBI TaxID	9606

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Cellosaurus	CVCL_0589

EXPRESSION AND SUSCEPTIBILITY

Antigen expression

CD4+ (37.1%, coexpressed with CD34 and CD33), CD13+(OKM13), CD15+(LeuM1), CD33+, CD34+(MY10), CD38+(OKT10, 50.1%), CD71+(Nu-TERf), HLA-DR+(OKDR).

HANDLING AND CULTURE

Culture medium

RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% heat-inactivated FBS

Subculturing

Maintain cultures by periodically adding or replacing the medium. Initiate cultures with a density of 5 x 10^5 cells/ml and keep the cell concentration within the range of 3 x 10^5 to 1 x 10^6 cells/ml for optimal growth.

Seeding density	1 x 10^5 cells/ml
Fluid renewal	Add fresh medium (20 to 30% by volume) every 2 to 3 days
Post-thaw recovery	About one week

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	7 years
Ethnicity	Japanese

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes