Lama-84

300261

GENERAL INFORMATION

Description

LAMA-84 is a human cell line derived from the peripheral blood of a patient with chronic myeloid leukemia (CML) in blast crisis. This cell line is characterized by the presence of the Philadelphia chromosome, which results in the BCR-ABL fusion gene, a hallmark of CML. The BCR-ABL oncogene is known for its role in increasing tyrosine kinase activity, which promotes various signaling pathways leading to uncontrolled cell proliferation and resistance to apoptosis. LAMA-84 cells are thus an invaluable model for studying the molecular mechanisms of CML progression and for evaluating the efficacy of tyrosine kinase inhibitors (TKIs) in a pre-clinical setting.
In research, LAMA-84 has been extensively used to understand the biology of CML, especially in the context of drug resistance and disease evolution. Studies involving this cell line have helped in elucidating the cellular responses to different generations of TKIs, such as imatinib, dasatinib, and nilotinib. Moreover, LAMA-84 has contributed to the investigation of new therapeutic strategies aimed at overcoming TKI resistance, including the testing of combination therapies that target other signaling pathways synergistically affected by the BCR-ABL fusion protein.

Tissue	Blood
Disease	Chronic myeloid leukemia

Synonyms

- LAMA-84
- LAMA84
- Lama84

CLASSIFICATION

Organism	Human
Sex	Female
Growth properties	Suspension
Biosafety level	1

CHARACTERISTICS

Morphology	Round cells
Doubling time	30 hours

IDENTIFIERS

Citation

Lama-84 (Cytion catalog number 300261)

NCBI TaxID	9606
Cellosaurus	CVCL_0388

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EXPRESSION AND SUSCEPTIBILITY

Surface antigens

GPIIb/IIIa+, GPIIIa+

Mutational profile

BCR-ABL1 pos

Viruses

EBNA, EA, and VCA were not detected

HANDLING AND CULTURE

Culture medium

RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% heat-inactivated FBS

Subculturing

Cells adhering to the bottom of the cell culture flask may be dislodged by shaking. Maintain cultures by periodically adding or replacing the medium. Initiate cultures with a density of 5 x 10^5 cells/ml and keep the cell concentration within the range of 3 x 10^5 to 1 x 10^6 cells/ml for optimal growth.

Seeding density	1 to 2 x 10^4 cells/cm^2
Post-thaw recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 hours.

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	29 years
Ethnicity	Caucasian

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes