# SK-UT-1

300455

## GENERAL INFORMATION

**Description**

[illegible] SK-UT-1 [illegible] (ULMS) [illegible] [illegible] [illegible] [illegible] SK-UT-1 [illegible] [illegible] [illegible] [illegible] [illegible] [illegible] [illegible] (CSCs) [illegible] [illegible] [illegible] CD133+ CSCs [illegible] SK-UT-1 [illegible] [illegible] [illegible]
[illegible] SK-UT-1 [illegible] CD133+ (CD133+ CSCs) [illegible] (tumorspheres) [illegible] [illegible] [illegible] (in vivo) [illegible] ($10^4$ [illegible]) [illegible] (xenograft models) [illegible] CD133+ [illegible] [illegible] [illegible] [illegible] [illegible] [illegible] (CSC) [illegible] [illegible] CD44, ALDH1 [illegible] BMI1 [illegible] CD133+ [illegible] CD133− [illegible]
[illegible] SK-UT-1 [illegible] ULMS [illegible] [illegible] [illegible] CD133+ [illegible] [illegible] ULMS [illegible]

| | |
|---|---|
| **Tissue** | [illegible] |
| **Disease** | [illegible] (mesoderm) [illegible] leiomyosarcoma ([illegible] III) |

**Synonyms**

- SK UT 1
- SKUT-1
- SKUT1
- Skut1

## CLASSIFICATION

| | |
|---|---|
| **Organism** | Human |
| **Sex** | Female |
| **Growth properties** | Adherent |
| **Biosafety level** | 1 |

## CHARACTERISTICS

| | |
|---|---|
| **Morphology** | [illegible] |


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| | |
|---|---|
| **Tumourigenic** | □□□ □□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ |

**Karyotype**

(P8) □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□. □□□□□□□□□□□□□□□□□□□□□□: 0.0590

## IDENTIFIERS

**Citation**

SK-UT-1 (□□□□□□□□□□□□□□□□□□□ Cytion 300455)

| | |
|---|---|
| **NCBI TaxID** | 9606 |
| **Cellosaurus** | CVCL_0533 |

## EXPRESSION AND SUSCEPTIBILITY

**Isoenzymes**

Me-2, 1-2, PGM3, 1, PGM1, 1, ES-D, 1, AK-1, 1, GLO-1, 1-2, G6PD, B.

## HANDLING AND CULTURE

**Culture medium**

EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO3, w: EBSS (□□□□□□□□□□□□□□□ Cytion 820100a)

| | |
|---|---|
| **Incubation atmosphere** | 37°C, 5%CO2, □□□□□□□□□□□□□□ |

**Supplements**

□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ FBS 10% □□□ NEAA 1%

| | |
|---|---|
| **Dissociation reagent** | □□□□□□□□□ |

**Subculturing**

□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ PBS □□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□ □□□□□□□□□□□□ T25 □□□□□□□ PBS 3-5 □□. □□□□□□□□□□□□□□□ T75 □□□□□□□ 5-10 □□. □□□□□□□□□□□□□□□□□□ □□□□□□□□□□□ Accutase □□□□□□□ 1-2 □□. □□□□□□□□□□□□ T25 □□□ 2.5 □□. □□□□□□□□□□□□ T75□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□ 8-10 □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□ □ □□□□□□□□□□□□□□□□□□□□□□□ 10 □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□ 300 □□□□□□□□□□□□□□□□□□□□□□□□□□□□ (300xg) □□□□□□□□□□ 3 □□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□

| | |
|---|---|
| **Seeding density** | 1 x 10^4 □□□□□□/□□.^² |
| **Fluid renewal** | 2 □□□□□□□□□□□□□□□□□□□□ |

**Thawing and culturing**

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

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## QUALITY CONTROL AND CRYOPRESERVATION

### Sterility

[illegible] PCR [illegible]

[illegible] [illegible] [illegible] [illegible]

| | |
|---|---|
| **Freeze medium** | [illegible] [illegible] ([illegible] FBS) + 10% DMSO [illegible] [illegible] CM-1 ([illegible] Cytion 800100) [illegible] |

### Storage conditions

[illegible] [illegible] −150 [illegible] −196 °C [illegible] −80 °C [illegible]

### Shipping conditions

[illegible] [illegible] −78 °C [illegible] [illegible] [illegible]

## DONOR

| | |
|---|---|
| **Donor age** | 75 [illegible] |
| **Ethnicity** | [illegible] |

## LEGAL AND SAFETY

### Safety precautions

[illegible] [illegible] [illegible] [illegible] [illegible] [illegible]

### Warranty

[illegible]Cytion [illegible]
[illegible]
60 [illegible]
[illegible]
[illegible] [illegible]/[illegible]
[illegible] [illegible]
<a href=""/Who-we-are/Warranty/"" hreflang=""x-default"">[illegible].

| | |
|---|---|
| **Material Transfer Agreement required** | Yes |