WERI-Rb-1

300632

GENERAL INFORMATION

Description

The WERI-Rb-1 cell line is derived from a retinoblastoma, a rare malignant tumor of the retina that typically manifests in early childhood. This cell line was established to provide a consistent and replicable model for the study of retinoblastoma biology, offering insights into the genetic, molecular, and cellular mechanisms underlying this form of cancer. WERI-Rb-1 cells are particularly valued in oncological research for their utility in investigating the pathophysiological processes and potential therapeutic targets for retinoblastoma.
WERI-Rb-1 cells exhibit characteristics typical of retinoblastoma, including the expression of neuronal markers and the capability to form cell aggregates resembling Flexner-Wintersteiner rosettes, a hallmark of retinoblastoma histology. These cells have been used extensively to study the role of oncogenes and tumor suppressor genes in cancer development, with a focus on the RB1 gene, mutations of which are pivotal in the etiology of retinoblastoma. Furthermore, WERI-Rb-1 serves as an important tool in the evaluation of chemotherapeutic agents and novel drug delivery systems aimed at improving treatment outcomes for retinoblastoma patients.

Tissue	Eye
Disease	Retinoblastoma

Applications

3D cell culture

Synonyms

- WERI-RB-1
- WERI-Rb 1
- WERI-Rb1
- WERI-RB1
- WERI Rb-1
- WERIRb1
- WERI
- Wills Eye Research Institute-Retinoblastoma-1

CLASSIFICATION

Organism	Human
Sex	Female
Growth properties	Suspension
Biosafety level	1

CHARACTERISTICS

Morphology	Round cells
Tumourigenic	Yes, in rabbits

Karyotype

Human pseudodiploid karyotype with 3.9% polyploidy - 46(41-48)xx, +6, -10, -10, -14, -22, +3mar, add(3)(q25), add(3)(q25), add(4)(p15), add(5)(q35), i(6q), del(7)(p21), add(9)(q33), der(13)x2, add(16)(q23), add(16)(q23), i(17q), add(19)(q13) - apparently (uniparental?) disomic rearrangement of ch 13 - corresponds to reported karyotype

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IDENTIFIERS

Citation

WERI-Rb-1 (Cytion catalog number 300632)

NCBI TaxID	9606
Cellosaurus	CVCL_1792

EXPRESSION AND SUSCEPTIBILITY

Isoenzymes

ES-D, 1, G6PD, B, GLO-I, 2, Me-2, 1, PGM1, 1, PGM3, 0

Viruses

EBV -, HBV -, HCV -, HHV-8 -, HIV-1 -, HIV-2 -, HTLV-1/2 -, MLV -, SMRV -

Reverse transcriptase	Negative

HANDLING AND CULTURE

Culture medium

RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% FBS and 0.01 mg/mL insulin

Subculturing

Gently homogenize the cell suspension in the flask by pipetting up and down, then take a representative sample to determine the cell density per ml. Dilute the suspension to achieve a cell concentration of 1 x 10^5 cells/ml with fresh culture medium, and aliquot the adjusted suspension into new flasks for further cultivation.

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use 50% basal medium + 40% FBS + 10% DMSO, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	1 year

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes

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