Human Gingival Fibroblasts (hGF)

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- HGF
- HFIB-G
- Human fibroblasts
- gingiva
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□□□	Spindle-shaped (fibroblast-like)
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IBAN: DE35 6725 0020 0009 2076 86
BIC: SOLADES1HDB

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DMEM:□□ F12(1:1), w: 3.1 g/L □□□, w: 2.5mM L-□□□□, w: 15mM HEPES, w: 0.5mM □□□□ □□□, w: 1.2 g/L NaHCO3 (Cytion □□ □□ 820400a)

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□□□ 10% FBS, 10ng/mL bFGF, 10□□□□□□/L □□□□ □□□□□

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□□ □□	More than 4,000 cells/cm²
□□ □□	24 hours after seeding, then every 48 to 72 hours
□□ □ □□	Post-thaw viability ≥ 70 % viable cells. More than 80 % confluence within the expected recovery period.

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Thaw the cryovial quickly in a 37 °C water bath, swirling gently. Just before the last ice has melted, move the vial to a laminar flow hood and decontaminate its outer surface.
Transfer the cell suspension into a 15 ml centrifuge tube, add 10 ml of Fibroblast Growth Medium and centrifuge at 250 × g for 5 minutes.
Remove the supernatant and resuspend the cell pellet in 10 ml of Fibroblast Growth Medium. Check cell number and viability.
Divide the suspension between two 25 cm² culture flasks (5 ml each). Using two flasks reduces the risk of losing all cells if a passage fails.
Incubate at 37 °C, 5 % CO_2 and saturated humidity, with the screw caps opened by half a turn for gas exchange.
Change the medium after 24 hours to remove unattached cells, then every 48 to 72 hours, using pre-warmed medium only.
Subculture the cells at about 75 % confluence. Denser cultures undergo irreversible contact inhibition and stop proliferating. Use the subculture reagents recommended on the certificate of analysis.
Recommended seeding density: more than 4,000 cells/cm².

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