HROC24 T1 M1

300813

GENERAL INFORMATION

Description

□□ PD Dr. Michael Linnebacher □□□ PDTx (Patient-derived Tumor xenografts) □□□□□□□□□ 2006

Tissue	□□□□□□□□□□□, □□□□□□□□□□□ PDx (□□□□□□□□□□□□□□□□□□□□□□□□□□□□□) □□□ □□□□□□□□□□□ CRC □□□□ (□□□□□□□□□□□□□□□□□□□, UICC I, TNM stage T2N0M0R0L0V1 grading G2). Lk(n) + 0, Σ Lk(n) 13
Disease	□□□□□□□□□□□
Metastatic site	□□□□□□□□□□□□□□□□□ (UICC stage I; TNM T2N0M0; □□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□)

Applications

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Synonyms

- HROC24x

CLASSIFICATION

Organism	Human
Sex	Male
Growth properties	Adherent
GMO status	Non-GMO

GMO status details

No genetic modification; wildtype patient-derived MSI-H CRC cell line established from a patient-derived xenograft by PD Dr. Linnebacher

Biosafety level	1

CHARACTERISTICS

Morphology	□□□□□□□□□□□□□□□□□□□□□□□□□□□□
Cell type	□□□□□□□□□□□□□□□□□□
Tumourigenic	□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□
Ploidy status	□□□□□□□□□□
MSI status	MSI-H
Doubling time	20 □□□□□□□□

IDENTIFIERS

Citation

HROC24 T1 M1 ([illegible] Cytion 300813)

NCBI TaxID	9606
Cellosaurus	CVCL_1U80

EXPRESSION AND SUSCEPTIBILITY

Mutational profile

APCmut, p53wt, K-Raswt, N-Raswt, H-Ras wt, B-RAFV600E, PIK3CAwt

Viruses

[illegible] SV40, JC/BK, HBV, HCV, HIV

HANDLING AND CULTURE

Culture medium

DMEM:Ham's F12 (1:1), w: 3.1 [illegible]/[illegible] [illegible], w: 2.5 [illegible] L-[illegible], w: 15 [illegible] HEPES, w: 0.5 [illegible] [illegible], w: 1.2 [illegible]/[illegible] $NaHCO_3$ ([illegible] Cytion 820400a)

Incubation atmosphere	37°C, 5%CO_2, [illegible]

Supplements

[illegible] FBS 10%

Dissociation reagent	[illegible]

Subculturing

[illegible] PBS [illegible] [illegible] T25 [illegible] PBS 3-5 [illegible]. [illegible] T75 [illegible] 5-10 [illegible]. [illegible] Accutase [illegible] 1-2 [illegible]. [illegible] T25 [illegible] 2.5 [illegible]. [illegible] T75[illegible] 8-10 [illegible] 10 [illegible] 300 [illegible] (300xg) [illegible] 3 [illegible]

Split ratio	1 [illegible] 3
Seeding density	2 x 10^4 [illegible]/[illegible].^²
Fluid renewal	[illegible] 3 [illegible] 5 [illegible]
Post-thaw recovery	[illegible]

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

[illegible] PCR [illegible]

[illegible]

Freeze medium	[illegible] ([illegible] FBS) + 10% DMSO [illegible] CM-1 ([illegible] Cytion 800100) [illegible]

Storage conditions

[illegible] −150 [illegible] −196 °C [illegible] −80 °C [illegible]

Shipping conditions

[illegible] −78 °C [illegible]

DONOR

Donor age	98 [illegible]
Ethnicity	[illegible]

LEGAL AND SAFETY

Safety precautions

[illegible]

Warranty

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Third-party agreements apply	Yes

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