Daudi

302009

GENERAL INFORMATION

Description

[illegible] Daudi [illegible] 1967 [illegible] 16 [illegible] Burkitt [illegible] Daudi [illegible] Epstein-Barr (EBV) [illegible] Burkitt [illegible] EBV [illegible]

[illegible] Daudi [illegible] Major Histocompatibility Complex (MHC) class I [illegible]-2-[illegible] MHC class I [illegible]-2-[illegible] Daudi [illegible]

[illegible] Daudi [illegible]

[illegible] Daudi [illegible]

Tissue	[illegible]
Disease	[illegible]

Applications

[illegible] B, [illegible], [illegible], [illegible], [illegible].

Synonyms

- DAUDI
- NK-10A
- NK-10a
- NK 10a
- NK10a
- N
- GM03190
- GM3190
- GM03190A
- GM17346

CLASSIFICATION

Organism	Human
Sex	Male
Growth properties	Suspension

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Banking information
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BIC: SOLADES1HDB

CHARACTERISTICS

Morphology	เซลล์กลม
Cell type	ลิมโฟบลาสต์ชนิดบี

Karyotype

46 เกือบเป็นแบบไดพลอยด์

IDENTIFIERS

Citation

ดาวดี (หมายเลขแคตตาล็อก Cytion 302009)

NCBI TaxID	9606
Cellosaurus	CVCL_0008

EXPRESSION AND SUSCEPTIBILITY

Antigen expression

CD10+, CD19+, CD20+, CD21+, CD22+, CD23-, CD24-, CD32+, CD37+, CD38+, CD39-,CD40+, CD54+, CD72+, CD73-, CD75+, CD77+, CD81+, CD82+, CD83-, CD84+, CD86+

HANDLING AND CULTURE

Culture medium

RPMI 1640, w: 2.0 mM กลูตามีนเสถียร, w: 2.0 กรัม/ลิตร NaHCO3 (หมายเลขบทความ Cytion 820700a)

Incubation atmosphere	37°C, 5%CO2, บรรยากาศชื้น

Supplements

เติมอาหารเลี้ยงเชื้อด้วย FBS ที่ทำให้ไม่ทำงานทางชีวภาพด้วยความร้อน 10%

Subculturing

รักษาวัฒนธรรมโดยการเติมหรือเปลี่ยนอาหารเลี้ยงเชื้อเป็นระยะๆ เริ่มต้นวัฒนธรรมด้วยความหนาแน่น 5×10^5 เซลล์/มล . และรักษาความเข้มข้นของเซลล์ให้อยู่ในช่วง 3×10^5 ถึง 1×10^6 เซลล์/มล . เพื่อการเจริญเติบโตที่ดีที่สุด

Seeding density	3×10^5 เซลล์/มิลลิลิตร
Fluid renewal	2 ครั้งต่อสัปดาห์
Post-thaw recovery	รวดเร็ว (48 ชั่วโมง)

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.

Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.

For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.

Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.

Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.

Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.

Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.

Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

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QUALITY CONTROL AND CRYOPRESERVATION

Sterility

[illegible] PCR [illegible]

[illegible]

Freeze medium	[illegible] ([illegible] FBS) + 10% DMSO [illegible] CM-1 ([illegible] Cytion 800100) [illegible]

Storage conditions

[illegible] −150 [illegible] −196 °C [illegible] −80 °C [illegible]

Shipping conditions

[illegible] −78 °C [illegible]

DONOR

Donor age	16 [illegible]
Ethnicity	[illegible]

LEGAL AND SAFETY

Safety precautions

[illegible]

Warranty

[illegible]Cytion [illegible]
[illegible]
60 [illegible]
[illegible]
[illegible] [illegible]/[illegible]
[illegible]
<a href=""/Who-we-are/Warranty/"" hreflang=""x-default"">[illegible].

Material Transfer Agreement required	Yes

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