35.1

305002

GENERAL INFORMATION

Description

35.1 □□□□□□□□Mus musculus□B □□□□□□□□□□□□□□□□□ T □□□□□□ Tp50□CD2□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ 35.1 □□□□□□□□□□□ T □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ 35.1 □□□□□□□□□ NSI/1 □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ CD2 □□□□□□□□□□□□□□□□ IgG2a□

Tissue	□□□B □□□□□□□□□NSI/1 □□□□□□□□□□□□□□
Disease	□□□□B □□ × □□□□□□□□□□□□□□□□□□ CD2 □□□□□□□Tp50□
Metastatic site	□□□□□□□□□□□□□□□□□□□□□□

Applications

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Synonyms

- HB-222

CLASSIFICATION

Organism	Mouse
Sex	Unspecified
Growth properties	Suspension
GMO status	GMO-S1

GMO status details

GMO-S1: This hybridoma contains rearranged immunoglobulin genes from a B cell-myeloma fusion. IgG2a heavy and light chain genes are expressed from the B cell fusion partner. This classification applies only within Germany and may differ elsewhere.

Biosafety level	2

CHARACTERISTICS

Morphology	□□□□□
Cell type	B □□□□□□□□□□□□
Doubling time	□ 24 □ 36 □□

IDENTIFIERS

Citation

1

NCBI TaxID	10090
Cellosaurus	CVCL_7239

EXPRESSION AND SUSCEPTIBILITY

Protein expression

□□□□ CD2 □□□□□□□□□□□□□

HANDLING AND CULTURE

Culture medium

RPMI 1640□□ 2.0 mM □□□□□□□□ 2.0 g/L $NaHCO_3$□Cytion □□□□ 820700a□

Incubation atmosphere	37°C□5% CO_2□□□□□□

Supplements

□□□□□□□ 10% FBS □ 0.05 mM 2-□□□□

Dissociation reagent	□

Subculturing

□□□ 1 × 10^5 □□□/□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□

Split ratio	1 □ 5
Seeding density	1 □ 5 × 10^5 □□□/□□
Fluid renewal	□□ 2 □ 3 □
Post-thaw recovery	□□□□□□□□ 300×g □□ 5 □□□□□□□□□□□□□□□□□□□□ 1 × 10^5 □□/ml □□□□□□□□□□□ □□□□□□□□□□□□ □□□□□□□□□□□□□□□□□ 24–48 □□□□□□□□□□□□□□

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

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Freeze medium	□□□□□□□□□□□□□□□□□□□□□□□□□ FBS□+ 10% DMSO □□□□□□□□□□□□□□□ CM-1 □Cytion □□ 800100□□ □□

Storage conditions

□□□□□□□□□□□□□□□□□ −150 □ −196 °C □□□□□□□□□□□□□□□□□□□□□□□□□□□□□ −80 °C □□□□□□□□□□□□□□

Shipping conditions

□□ −78 °C□□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□

DONOR

Breed / subspecies	BALB/c□NSI/1 □□□□□□□□□
Donor age	□□□□
Ethnicity	□□□□□□□□□□

LEGAL AND SAFETY

Safety precautions

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Warranty

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Material Transfer Agreement required	Yes

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