TM3

305167

GENERAL INFORMATION

Description

TM3 □□□□□□□ 11 □ 13 □□ □□□□□□□□ F2a □□□□□□□□□□□□□□□□□□□□□□□□□□LH□□□□□□□□□□□EGF□□□□□□□□□□□□□□□□□□□□□□□□□□□□ TM3 □□□□□□□□□□□□□ LH □□□□□□□□□ cAMP □□□□□□□□□□□□□□□□□□□ (FSH) □□□□□LH □□□□□□□□□□□□□□□□□□ □□□□LH□□

Tissue	□□
Disease	□□□□□□□□□□□□□□□□□BALB/c □□□
Metastatic site	□□□□□□□□□□□□□□□□□□□□

Applications

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Synonyms

- TM-3

CLASSIFICATION

Organism	Mouse
Sex	Male
Growth properties	Adherent
GMO status	Non-GMO

GMO status details

No genetic modification; wildtype mouse Leydig cell line derived from neonatal BALB/c testis by primary culture

Biosafety level	1

CHARACTERISTICS

Morphology	□□□
Cell type	□□□□□□
Doubling time	□ 36 □ 48 □□

IDENTIFIERS

Citation

TM3□Cytion □□ 305167□

NCBI TaxID	10090
Cellosaurus	CVCL_4326

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Local court Mannheim - HRB 714394
CEO / Geschäftsführer: Jonathan Steubing

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Sparkasse Heidelberg
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BIC: SOLADES1HDB

HANDLING AND CULTURE

Culture medium

DMEM□Ham's F12□1:1□□□□□□3.1 g/L □□□□□□□□2.5 mM L-□□□□□□□□□15 mM HEPES□ w: 0.5 mM □□□□□w: 1.2 g/L $NaHCO_3$ □Cytion □□□□ 820400a□

Incubation atmosphere	37°C□5% CO_2□□□□□□

Supplements

□□□□□□□ 2.5% □□□□□FBS□□ 5% □□□

Dissociation reagent	Accutase

Subculturing

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Split ratio	1 □ 3
Seeding density	1 □ 3 × 10^4 □□□/cm²
Fluid renewal	□□ 2 □ 3 □
Post-thaw recovery	□□□□□□□□ 5 × 10^4 □□□/cm² □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ 24–48 □ □□□□□□□□ FBS □ LH □ cAMP □□□□□□□□□□□□□□□□ LH □□□□

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

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Freeze medium	□□□□□□□□□□□□□□□□□□□□□□□□□ FBS□+ 10% DMSO □□□□□□□□□□□□□□□ CM-1 □Cytion □□ 800100□□ □□

Storage conditions

□□□□□□□□□□□□□□□□□□□ −150 □ −196 °C □□□□□□□□□□□□□□□□□□□□□□□□□□□□□ −80 °C □□□□□□□□□□□□□□□

Shipping conditions

□□ −78 °C□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□

DONOR

Breed / subspecies	BALB/c
Donor age	11 □ 13 □

LEGAL AND SAFETY

Safety precautions

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Warranty

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Material Transfer Agreement required	Yes

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