H4-II-E-C3

305336

GENERAL INFORMATION

Description

The H4-II-E-C3 cell line is a well-established rat hepatoma cell line derived from liver carcinoma. This line is widely used in research focused on liver-specific functions and the metabolic processes that occur within hepatocytes. One notable aspect of H4-II-E-C3 cells is their high expression of the high-affinity glutamate transporter GLT-1A. This feature distinguishes them from normal rat hepatocytes, which do not exhibit significant levels of this transporter. The expression of GLT-1A in H4-II-E-C3 cells allows for the efficient study of glutamate metabolism and transport, providing insights into liver cell transformation and tumor cell metabolism.
Additionally, H4-II-E-C3 cells are used to study aldehyde metabolism, particularly the activity of mitochondrial class 2 aldehyde dehydrogenase (ALDH2). This enzyme is critical for metabolizing acetaldehyde produced during ethanol oxidation, making this cell line suitable for research on alcohol metabolism and its cellular effects. The use of an "acetaldehyde clamp" technique in these cells has allowed precise measurement of ALDH2 activity without the need for mitochondrial isolation, facilitating the study of enzyme kinetics and the impact of various inhibitors on aldehyde detoxification.

Tissue	Liver
Disease	Hepatocellular carcinoma

Synonyms

- H4IIEC3
- H4-IIE-C3
- H4-II-EC3
- H-II-E-C3

CLASSIFICATION

Organism	Rat
Sex	Male
Growth properties	Adherent
GMO status	GMO-S1

GMO status details

GMO-S1: This rat hepatocellular carcinoma cell line (H4-II-E-C3) contains undefined modifications classified as GMO, stably present in the transformed hepatic cell line. This classification applies only within Germany and may differ elsewhere.

Biosafety level	1

CHARACTERISTICS

Tumourigenic	Yes, in AxC rats

IDENTIFIERS

Citation

H4-II-E-C3 (Cytion catalog number 305336)

NCBI TaxID	10116

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Cellosaurus	CVCL_0285

EXPRESSION AND SUSCEPTIBILITY

Protein expression

Gene expression: tyrosine aminotransferase, albumin, transferrin, prothrombin

HANDLING AND CULTURE

Culture medium

DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% FBS

Dissociation reagent	Accutase

Subculturing

Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

Seeding density	1-3*10^4/cm²
Fluid renewal	2 to 3 times per week

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Breed / subspecies	AxC
Donor age	Unspecified

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes