ZR-75-30

305389

GENERAL INFORMATION

Description

ZR-75-30 is a human breast cancer cell line derived from a ductal carcinoma. Genomic profiling studies have shown that ZR-75-30 harbors amplification of the ERBB2/HER2 gene, a key driver in a subset of breast cancers. This amplification results in elevated HER2 protein expression, which has been linked to increased proliferation and resistance to certain therapies. Additionally, ZR-75-30 exhibits alterations in the epidermal growth factor receptor (EGFR) signaling pathway, including gains of EGFR-related genes, suggesting that the cell line may be useful in studying HER2-targeted therapies and their resistance mechanisms.
Transcriptomic analyses have placed ZR-75-30 within the luminal subtype of breast cancer, supporting its relevance for studying endocrine therapy responses. The cell line has been included in studies evaluating precision medicine approaches, where molecular profiling has helped predict responses to targeted treatments. Given its molecular characteristics, ZR-75-30 is widely used as a preclinical model for evaluating hormone receptor-targeted therapies and HER2 inhibitors, making it a valuable tool in breast cancer research.

Tissue	Breast, Mammary gland
Disease	Invasive breast carcinoma of no special type
Metastatic site	Ascites

Synonyms

- ZR75-30
- ZR7530

CLASSIFICATION

Organism	Human
Sex	Female
Growth properties	Adherent
Biosafety level	1

CHARACTERISTICS

Morphology	Epithelial
Cell type	Epithelial
Doubling time	110 hours

IDENTIFIERS

Citation

ZR-75-30 (Cytion catalog number 305389)

NCBI TaxID	9606
Cellosaurus	CVCL_1661

EXPRESSION AND SUSCEPTIBILITY

Mutational profile

Mutation: Gene fusion, APPBP2 + HGNC, PHF20L1, Name(s)=APPBP2-PHF20L1.Gene fusion, BCAS3 + HGNC, HOXB9, Name(s)=BCAS3-HOXB9. Gene fusion, COL14A1 + HGNC, SKAP1, Name(s)=COL14A1-SKAP1. Gene fusion, DDX5 + HGNC, DEPTOR, Name(s)=DDX5-DEPTOR. Gene fusion, BCAS3 + HGNC, ERBB2, Name(s)=ERBB2-BCAS3. Gene fusion, ENPP2 + HGNC, PLEC, Name(s)=PLEC-ENPP2, PLEC1-ENPP2. Gene fusion, PCGF2 + HGNC, TAOK1, Name(s)=TAOK1-PCGF2. Gene fusion, NRIP1 + HGNC, TIAM1, Name(s)=TIAM1-NRIP1. Gene fusion, ARHGAP32 + HGNC, TIMM23, Name(s)=TIMM23-ARHGAP32. Gene fusion, LASP1 + HGNC, TRPS1, Name(s)=TRPS1-LASP1. Gene fusion, CWC25 + HGNC, USP32, Name(s)=USP32-CWC25, USP32-CCDC49. Gene fusion, OPRD1 + HGNC, ZMYM4, Name(s)=ZMYM4-OPRD1. Mutation, BRAF, Simple, p.Ile326Thr (c.977T>C), Heterozygous, CDH1, Simple, p.Glu243Ter (c.727G>T), Homozygous.

HANDLING AND CULTURE

Culture medium

RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% FBS, 10 µg/ml Insulin

Fluid renewal	2 to 3 times per week

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	47 years
Ethnicity	African American

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes