JIMT-1

305433

GENERAL INFORMATION

Description

The JIMT-1 cell line is derived from a HER2-positive human breast carcinoma and is known for its resistance to trastuzumab, a commonly used HER2-targeted therapy. This makes JIMT-1 a valuable model for studying mechanisms of resistance to anti-HER2 treatments and for developing new therapeutic strategies. Unlike many other HER2-positive breast cancer cell lines, JIMT-1 mimics clinical cases where initial responses to HER2-targeted therapies are observed, but resistance subsequently develops. This feature has made it instrumental in exploring the efficacy of new drugs and combination therapies aimed at overcoming trastuzumab resistance.
JIMT-1 cells are also employed in studies investigating the interplay between HER2 and other signaling pathways, such as those involving the epidermal growth factor receptor (EGFR). Cross-talk between these pathways contributes to the cells' resistance to conventional therapies.
Research has shown that JIMT-1 cells respond variably to different tyrosine kinase inhibitors (TKIs) and antibody-drug conjugates (ADCs). For example, while the cell line exhibits resistance to trastuzumab-emtansine (T-DM1) and shows only partial sensitivity to newer agents like trastuzumab-deruxtecan (T-DXd), it has been demonstrated that alternative ADCs such as disitamab vedotin (DV) might offer enhanced efficacy.
In vitro studies highlight the versatility of JIMT-1 for screening drugs that target not only HER2 but also other molecular pathways. These studies provide critical data for evaluating the synergistic effects of combination treatments involving ADCs and TKIs or novel targeted therapies. The cell line's behavior in various drug resistance scenarios underscores its importance in preclinical drug development, particularly for HER2-positive breast cancer with acquired or intrinsic resistance.

Tissue	Breast
Disease	Breast ductal carcinoma
Metastatic site	Pleural effusion

Synonyms

- JIMT1
- JIMT

CLASSIFICATION

Organism	Human
Sex	Female
Growth properties	Adherent
Biosafety level	1

CHARACTERISTICS

Morphology	Epithelial-like

IDENTIFIERS

Citation

JIMT-1 (Cytion catalog number 305433)

NCBI TaxID	9606
Cellosaurus	CVCL_2077

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EXPRESSION AND SUSCEPTIBILITY

Oncogenes

HER-2 (insensitive to HER-2-inhibiting drugs, e.g. trastuzumab), ER-, PR-, AR-

Mutational profile

Mutation: PIK3CA, p.Cys420Arg (c.1258T>C), heterozygous; Mutation: TP53, p.Arg248Trp (c.742C>T), homozygous

HANDLING AND CULTURE

Culture medium

DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% heat-inactivated FBS

Dissociation reagent	Accutase

Subculturing

Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

Seeding density	1 x 10^4 cells/cm^2

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	62 years
Ethnicity	Caucasian

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes