Huh-7.5.1

305491

GENERAL INFORMATION

Description

The Huh-7.5.1 cell line is a subclone derived from the Huh-7 hepatoma cells, known for its enhanced permissiveness to hepatitis C virus (HCV) replication. Developed as an improved version of the Huh-7.5 cell line, Huh-7.5.1 cells were established from a cured HCV replicon cell line and exhibit higher susceptibility to HCV infection compared to their predecessors. They are capable of supporting robust HCV replication cycles, which makes them a favored model for studying HCV life cycles, including viral entry, RNA replication, and the production of infectious particles.
The use of Huh-7.5.1 cells has been particularly valuable in HCV research due to their increased efficiency in viral RNA replication and infectious virus production. This cell line exhibits high viral titers, typically reaching 10^4 to 10^5 infectious units per milliliter in culture supernatants during HCV-JFH1 strain infections. This characteristic permits detailed analysis of viral pathogenesis and antiviral responses in vitro. Importantly, Huh-7.5.1 cells are employed in screening for antiviral compounds that target HCV, aiding the development of treatments against HCV and providing insights into resistance mechanisms.
Additionally, the cells have been used in broader studies involving viral interactions with host cell machinery, lipid metabolism, and related therapeutic targets. For example, inhibition of cellular pathways such as the SKI-1/S1P protease-mediated SREBP pathway has demonstrated impacts on virus replication within Huh-7.5.1 cells, highlighting their utility for research into host-directed antiviral strategies.

Tissue	Liver
Disease	Adult hepatocellular carcinoma

Synonyms

- Huh 7.5.1
- Huh7.5.1

CLASSIFICATION

Organism	Human
Sex	Male
Growth properties	Adherent
Biosafety level	1

IDENTIFIERS

Citation

Huh-7.5.1 (Cytion catalog number 305491)

NCBI TaxID	9606
Cellosaurus	CVCL_E049

EXPRESSION AND SUSCEPTIBILITY

Mutational profile

Mutation: KDR, p.Gln472His (c.1416A>T), homozygous; Mutation: POLD3, p.Lys109Arg (c.326A>G) (p.Lys70Arg, c.209A>G); Mutation: RIGI, p.Thr55Ile (c.164C>T); Mutation: TERT, c.1-124C>T (c.228C>T) (C228T); Mutation: TP53, p.Tyr220Cys (c.659A>G), homozygous

Virus susceptibility

Compared to the parent Huh-7 cell line, permits improved hepatitis C virus (HCV) production when infected with strain JFH-1.

Cytion GmbH
Nikola-Tesla-Str. 3
69124 Heidelberg
Tel: +49 (0)6221 405780
info@cytion.com
cytion.com

Registered office Heidelberg
VAT ID: DE283292917
Local court Mannheim - HRB 714394
CEO / Geschäftsführer: Jonathan Steubing

Banking information
Sparkasse Heidelberg
IBAN: DE35 6725 0020 0009 2076 86
BIC: SOLADES1HDB

HANDLING AND CULTURE

Culture medium

DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% FBS

Dissociation reagent	Accutase
Seeding density	2-4*10^4/cm²

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	57 years
Ethnicity	Japanese

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes

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