SNB-19

305492

GENERAL INFORMATION

Description

The SNB-19 cell line is a human glioblastoma multiforme (GBM) model derived from a high-grade glioma tumor. It is one of the widely studied glioma cell lines and is used for exploring the biology of aggressive brain tumors, especially glioblastoma. SNB-19 cells exhibit epithelial morphology and are adherent in culture. They have been extensively used in studies of tumor proliferation, invasion, and response to therapy, particularly for investigating glioblastoma's resistance mechanisms to conventional treatments.
Genomic profiling of SNB-19 cells has revealed important genetic alterations commonly associated with GBM, including mutations in tumor suppressor genes and oncogenes such as TP53, EGFR, and PTEN. These cells also demonstrate chromosomal abnormalities, including amplification of oncogenic drivers and deletions in tumor suppressor loci. The genetic landscape of SNB-19 provides an important model for studying the molecular pathways driving GBM pathogenesis and for identifying potential targets for therapy.
SNB-19 has been used extensively to evaluate the efficacy of novel chemotherapeutics and targeted agents. The cell line is also employed in assays studying glioblastoma's invasive and migratory properties, as it effectively mimics the highly invasive nature of GBM in vitro. Moreover, proteomic analyses of SNB-19 have contributed to understanding protein-level dysregulations and their correlation with genetic alterations in glioblastoma. These characteristics make SNB-19 an essential tool in translational research focused on glioblastoma.

Tissue	Brain, parietal lobe
Disease	Astrocytoma

Synonyms

- SNB.19
- SNB19
- Surgical Neurology Branch-19

CLASSIFICATION

Organism	Human
Sex	Male
Growth properties	Adherent
Biosafety level	1

CHARACTERISTICS

Morphology	Fibroblast-like
Cell type	Fibroblast
Doubling time	24 hours

IDENTIFIERS

Citation

SNB-19 (Cytion catalog number 305492)

NCBI TaxID	9606
Cellosaurus	CVCL_0535

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EXPRESSION AND SUSCEPTIBILITY

Mutational profile

Mutation: PTEN, Simple, p.Glu242Valfs*15 (c.723_724dupTG), Homozygous; Mutation: TERT, Simple, c.1-124C>T (c.228C>T) (C228T), Unspecified; Mutation: TP53, Simple, p.Arg273His (c.818G>A), Homozygous

HANDLING AND CULTURE

Culture medium

DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% FBS

Seeding density	1-4 x 10^4 cells/cm2
Fluid renewal	2 to 3 times per week

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	75 years
Ethnicity	Caucasian

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes

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