C6-Luc

305620

GENERAL INFORMATION

Description

C6-Luc □□□ C6 □□□□□□□□□□□□□□□□□□□□□□□□□ Benda □□□1968□□□□ Wistar □□□ N-□□□□□□□□□□□□□□□□□□□□□□ □□□ C6 □ □□ B □□□□□□□□□□□□□□□□□□□□□□□□□□

C6-Luc □□□BLI□□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□ □□□□□□□□□□□□□□ Wistar □ Sprague-Dawley □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□

Tissue	□□
Disease	□□□□□□□□

Applications

□□□

Synonyms

- Luciferase Reporter C6 Cell Line

CLASSIFICATION

Organism	Rat
Sex	Male
Growth properties	Adherent
GMO status	GMO-S1

GMO status details

GMO-S1: This cell line contains a stably integrated firefly luciferase reporter cassette (Luc2, codon-optimized) introduced via replication-incompetent lentiviral transduction. The resulting polyclonal cell population was maintained under puromycin selection (1–5 µg/mL). S1 containment is required. This classification applies only within Germany and may differ elsewhere.

Biosafety level	1

CHARACTERISTICS

Morphology	□□□□□□□□
Tumourigenic	□□□□□□□□□□ Wistar/Sprague-Dawley □□□□□□□□□□□□
Doubling time	24□48□□

IDENTIFIERS

Citation

C6-Luc□Cytion □□ 305620□

NCBI TaxID	10116
Cellosaurus	CVCL_E3GZ

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Registered office Heidelberg
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Local court Mannheim - HRB 714394
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Banking information
Sparkasse Heidelberg
IBAN: DE35 6725 0020 0009 2076 86
BIC: SOLADES1HDB

EXPRESSION AND SUSCEPTIBILITY

Antigen expression

Luc2□□□□□□□□□□□□

Products

□□□□□□□□luc2□pGL4.50□

HANDLING AND CULTURE

Culture medium

RPMI 1640□□ 2.1 mM □□□□□□□□□ 2.0 g/L $NaHCO_3$□Cytion □□□□ 820700a□

Incubation atmosphere	37°C□5% CO_2□□□□□□□

Supplements

10% □□□□

Dissociation reagent	Accutase

Subculturing

□□ 0.25% □□□□□/EDTA□□□□□□□□70–80%□□□□□□□□□ 1:3 □ 1:6 □□□□□□□□□

Seeding density	1-3×10^4/cm^2

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 200 x g for 5 minutes, carefully discard the supernatant containing freezing medium.
Follow the procedure described under Post-Thaw Recovery

QUALITY CONTROL AND CRYOPRESERVATION

Freeze medium	□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ 10% □□□□□□□□DMSO□□□□□□□□□□□□□□□□□□□□□□□□□□

Storage conditions

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Shipping conditions

□□ −78 °C□□

DONOR

Donor age	□□□□

LEGAL AND SAFETY

Safety precautions

□□ □□

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Warranty

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Material Transfer Agreement required	Yes