SNU-638

305634

GENERAL INFORMATION

Description

The SNU-638 cell line is a human gastric carcinoma model established from the ascitic fluid of a male gastric cancer patient. It exhibits poor differentiation and minimal desmoplasia, and in vitro it grows in a mixed pattern with heterogeneous density and poor attachment to the culture substrate. The cells maintain a round to oval contour and display a low nuclear-to-cytoplasmic ratio, with limited development of microvilli. These characteristics reflect features commonly associated with aggressive gastric cancer phenotypes and make the line suitable for studying poorly differentiated gastric adenocarcinomas.
At the molecular level, SNU-638 does not harbor mutations in the *c-Ki-ras* gene but does express high levels of tumor-associated markers such as CA 19-9 and tissue polypeptide antigen (TPA), with absent alpha-fetoprotein (AFP) expression. It also carries a *TP53* gene mutation, which is frequently found in gastric cancers and plays a central role in tumorigenesis. Genomic profiling revealed that SNU-638 lacks MET amplification or overexpression, categorizing it as MET-negative with minimal dependency on the MET signaling pathway. This molecular profile makes SNU-638 a valuable control cell line in studies targeting MET or evaluating the efficacy of MET inhibitors in gastric cancer.

Tissue	Gastric
Disease	Adenocarcinoma
Metastatic site	Ascites

Synonyms

- SNU638

CLASSIFICATION

Organism	Human
Sex	Male
Growth properties	Adherent
Biosafety level	1

CHARACTERISTICS

Morphology	Epithelial-like
Cell type	Epithelial
Doubling time	25 hours

IDENTIFIERS

Citation

SNU-638 (Cytion catalog number 305634)

NCBI TaxID	9606
Cellosaurus	CVCL_0102

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EXPRESSION AND SUSCEPTIBILITY

Mutational profile

Mutation: MET, Simple, p.Asn375Ser (c.1124A>G), Unspecified; Mutation: TP53, Simple, p.Arg282Trp (c.844C>T), Heterozygous

HANDLING AND CULTURE

Culture medium

RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% heat-inactivated FBS

Dissociation reagent	Accutase

Subculturing

Remove medium, add fresh 0.25 % trypsin 0.02 % EDTA solution, stand culture flask at 37""C for 3 to 5 minutes, add culture medium and collect the cells, transfer the medium into 15ml tube, centrifuge, aspirate the medium, resuspend the pellets with culture medium and dispense into the culture flask

Fluid renewal	2 to 3 times per week

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	48 years
Ethnicity	Korean

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes

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