ID8-Luc

305677

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ID8-Luc is a bioluminescent derivative of the murine ID8 ovarian surface epithelial cell line, engineered to stably express a firefly luciferase reporter gene. The parental ID8 cell line was derived from normal C57BL/6 mouse ovarian surface epithelial cells through serial in vitro passaging, during which the cells acquired a transformed, tumorigenic phenotype. ID8 cells grow as a peritoneal carcinomatosis-like model when injected intraperitoneally into syngeneic C57BL/6 mice, producing ascites and disseminated peritoneal tumors that closely recapitulate features of advanced human high-grade serous ovarian cancer. The model is widely used for studying ovarian tumor-immune interactions, tumor microenvironment biology, and immunotherapy approaches in an immunocompetent host.
The stable luciferase integration in ID8-Luc enables sensitive, noninvasive bioluminescence imaging (BLI) of tumor burden in living animals following administration of the luciferin substrate. The emitted signal correlates with viable tumor cell number, supporting longitudinal monitoring of peritoneal tumor dissemination, ascites development, and therapeutic response without repeated invasive procedures. This makes ID8-Luc especially valuable for preclinical studies evaluating immune checkpoint inhibitors, adoptive cell therapies, cancer vaccines, and novel targeted agents in a fully immunocompetent syngeneic model of ovarian cancer.
ID8-Luc retains the established tumorigenic and immunological properties of the parental ID8 line, including its intraperitoneal growth pattern and compatibility with C57BL/6 syngeneic hosts. The luciferase modification enhances experimental sensitivity and enables real-time assessment of disease progression and treatment efficacy. Researchers should validate luciferase activity, growth kinetics, and immunological phenotype under their specific experimental conditions prior to use in quantitative imaging or immunotherapy studies.

□□□□□	Ovary, surface epithelium
□□	Ovarian surface epithelial neoplasm

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- ID-8
- ID8/MOSEC

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GMO□□□	GMO-S1

GMO□□□□□□□□□□□□□

GMO-S1: This cell line contains a stably integrated firefly luciferase reporter cassette (Luc2, codon-optimized) introduced via replication-incompetent lentiviral transduction. The resulting polyclonal cell population was maintained under puromycin selection (1–5 µg/mL). S1 containment is required. This classification applies only within Germany and may differ elsewhere.

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□□□	Epithelial-like
□□□□□	Epithelial cell

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ID8-Luc (Cytion catalog number 305677)

NCBI □□□□□	10090
□□□□□□	CVCL_C8UV

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Luc2 (firefly, codon-optimized)

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DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)

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Supplement the medium with 10% FBS

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Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

□□□□	1 to 3
□□□□	2-4*10^4/cm²
□□□□□	2 to 3 times per week

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Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 200 x g for 5 minutes, carefully discard the supernatant containing freezing medium.
Follow the procedure described under Post-Thaw Recovery

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□□□□	As a cryopreservation medium, we use complete growth medium + 10% DMSO for adequate post-thaw viability.

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For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

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Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

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□□□□□□	Adult

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When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

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At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

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