K7M2 wt-Luc

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- K7M2-WT
- K7M2

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□□□ □□ □□	GMO-S1

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GMO-S1: This cell line contains a stably integrated firefly luciferase reporter cassette (Luc2, codon-optimized) introduced via replication-incompetent lentiviral transduction. The resulting polyclonal cell population was maintained under puromycin selection (1–5 µg/mL). S1 containment is required. This classification applies only within Germany and may differ elsewhere.

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K7M2 wt-Luc (Cytion □□ □□ 305684)

NCBI □□ □□□□	10090
□□□□□□	Not assigned (K7M2 wt-Luc reporter derivative; parental K7M2 syngeneic BALB/c osteosarcoma)

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□□(C3), □□, Fc □□□, IgG, □□□□ I(Fcgr1), □□

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Luc2 (□□□□, □□ □□□)

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DMEM, w: 4.5g/L □□□, w: 4mM L-□□□□, w: 3.7g/L NaHCO3, w: 1.0mM □□□□ □□□(□□□□ □□ □□ 820300a)

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10% FBS□ □□□ □□□□□

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□□ □□	1-3×10⁴/cm²
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Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 200 x g for 5 minutes, carefully discard the supernatant containing freezing medium.
Follow the procedure described under Post-Thaw Recovery

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