SW480-Luc

305698

GENERAL INFORMATION

Description

SW480-Luc □□□ SW480 □□ □□SW480□□□□□□□□51□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□ SW480 □□□□□□□□□□□□□□□□□□□□□□□□□□□ KRAS G12V□TP53 R273H □ P309S □□ APC □□□ □□□□□□□□□□□□□□ Wnt/β-catenin □ KRAS □□□□□□□□□□□□□□□□□□□□ □□□□□SW480 □ SW620 □□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□

SW480-Luc □□ (BLI)□ □□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ SW480-Luc □□□□□□□□□□ KRAS □□□□□Wnt □□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□ SW620 □□□□□□□□□□

SW480-Luc □□□□□ SW480 □□□□□□□□□□□□□□ KRAS□TP53 □ APC □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ □□□□□□□□ □□□

Tissue	□□
Disease	□□□□

CLASSIFICATION

Organism	Human
Sex	Male
Growth properties	Adherent
GMO status	GMO-S1

GMO status details

GMO-S1: This cell line contains a stably integrated firefly luciferase reporter cassette (Luc2, codon-optimized) introduced via replication-incompetent lentiviral transduction. The resulting polyclonal cell population was maintained under puromycin selection (1–5 µg/mL). S1 containment is required. This classification applies only within Germany and may differ elsewhere.

Biosafety level	1

CHARACTERISTICS

Morphology	□□□
Tumourigenic	□□□□□□□

IDENTIFIERS

Citation

SW480-Luc□Cytion □□□□ 305698□

NCBI TaxID	9606
Cellosaurus	CVCL_C8Y1

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Local court Mannheim - HRB 714394
CEO / Geschäftsführer: Jonathan Steubing

Banking information
Sparkasse Heidelberg
IBAN: DE35 6725 0020 0009 2076 86
BIC: SOLADES1HDB

EXPRESSION AND SUSCEPTIBILITY

Receptors expressed

□□□□□□□EGF□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□ Matrilysin □□□□□

Protein expression

P53□□□Luc2□□□□□□□□□□□□

Antigen expression

HLA A2□B8□B17□A□□□Rh+□Luc2□□□□□□□□□□□□

Isoenzymes

G6PD□B□PGM1□2□PGM3□1□6PGD□A□PEP-D□1□ES-D□1

Mutational profile

□□□p.Gln1338Ter□□□□□□□□p.Gly12Val□□□□□□□□p.Arg273His□□□□□□□□p.Pro309Ser□□□

Viruses

□□□□□□

Virus susceptibility

□□□□□□□□□□HIV□LAV□

Products

□□□□□CEA□0.7 □□/106 □□□/10 □□□□□□TGF-β□□□□□□□□□□□□□ GM-CSF□

HANDLING AND CULTURE

Culture medium

Ham's F12□w□1.0 mM □□□□□□□w□1.0 mM □□□□□w□1.1 g/L NaHCO3□Cytion □□□□ 820600a□□

Incubation atmosphere	37°C, 5%CO2, □□□□□

Supplements

□□□□□□□ 10% FBS

Dissociation reagent	Accutase

Subculturing

□□□□□□□□□□□□□□□□□□□□ PBS □□□T25 □□□ 3-5 □□ PBS□T75 □□□ 5-10 □□□□□□ Accutase □□□□□□□T25 □□□ 1-2 □□□T75 □□□ 2.5 □□□□□□□□□□□□ 8-10 □□□□□□□□□□□□□□ 10 □□□□□□□□□□□□□□□□□□□□ 300xg □□ 3 □□□□□□□□□□□□□□□□□□□□ □□□□□□□□□□□□□□□□□□□□

Split ratio	1 □ 3
Seeding density	1-3×10⁴/cm²
Fluid renewal	□□ 2 □ 3 □

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 200 x g for 5 minutes, carefully discard the supernatant containing freezing medium.
Follow the procedure described under Post-Thaw Recovery

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QUALITY CONTROL AND CRYOPRESERVATION

Freeze medium	□□□□□□□□□□□□□□□□□□□□□□+10% DMSO□□□□□□□□□□□□□□□

Storage conditions

□□□□□□□□□□□□□□□□□□□□□□□□□ -150 □ -196 ℃□□-80 °C□□□□□□□□□□□□□□□□□□□□

Shipping conditions

□□□ -78 °C □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□

DONOR

Donor age	51 □
Ethnicity	□□□□

LEGAL AND SAFETY

Safety precautions

□□

Warranty

□Cytion□□□□□□□□□□□□□□□
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Material Transfer Agreement required	Yes