NCI-H1755

305834

GENERAL INFORMATION

Description

NCI-H1755 is a human non-small cell lung cancer (NSCLC) cell line derived from a lung adenocarcinoma. It is part of the extensive National Cancer Institute (NCI) panel of thoracic cancer models, developed to support translational research into lung cancer biology and therapeutic response. This cell line exhibits a KRAS mutation, a feature common in many lung adenocarcinomas that contributes to constitutive activation of MAPK and PI3K signaling pathways, promoting uncontrolled cell growth and resistance to certain targeted therapies.
NCI-H1755 is included in several large-scale functional genomic and pharmacogenomic screens, including those profiling protein expression and response to targeted agents. Its molecular signature indicates activity in PI3K/AKT and RAS/RAF/MEK signaling pathways, which has made it a valuable tool for evaluating the effects of MEK inhibitors and other agents targeting downstream effector molecules. The cell line has also contributed to research focused on epithelial polarity, with studies identifying structural disruptions in polarity complex genes, such as PARD3, across various epithelial cancers including lung adenocarcinoma.
In vitro, NCI-H1755 cells grow in adherent monolayers and demonstrate epithelial morphology. They are maintained under standard culture conditions in RPMI-1640 medium supplemented with 10% fetal bovine serum. Due to its reproducible growth characteristics, mutational profile, and inclusion in molecular oncology datasets, NCI-H1755 is a frequently used model for investigating mechanisms of tumor progression, drug resistance, and potential therapeutic targets in KRAS-mutant NSCLC.

Tissue	Metastatic
Disease	Lung adenocarcinoma

Synonyms

- H1755
- H-1755
- NCIH1755

CLASSIFICATION

Organism	Human
Sex	Female
Growth properties	Adherent and suspension
Biosafety level	1

CHARACTERISTICS

Cell type	Epithelial-like and/or rounded

IDENTIFIERS

Citation

NCI-H1755 (Cytion catalog number 305834)

NCBI TaxID	9606
Cellosaurus	CVCL_1492

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EXPRESSION AND SUSCEPTIBILITY

Mutational profile

Mutation: BRAF, Simple, p.Gly469Ala (c.1406G>C), Heterozygous, TP53, Simple, p.Cys242Phe (c.725G>T), Homozygous

HANDLING AND CULTURE

Culture medium

RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

Incubation atmosphere	37°C, 5% CO_2, humidified atmosphere.

Supplements

Supplement the medium with 10% FBS

Dissociation reagent	Accutase
Fluid renewal	2 to 3 times per week

Thawing and culturing

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

QUALITY CONTROL AND CRYOPRESERVATION

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

DONOR

Donor age	65 years
Ethnicity	Caucasian

LEGAL AND SAFETY

Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

Disclaimer

Our cells are provided for in vitro laboratory research purposes exclusively and are not intended for clinical or diagnostic use, nor are they to be administered to humans or used for veterinary purposes. Users must adhere to all applicable guidelines and regulations for the handling and use of these cells in a research setting.

Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

Material Transfer Agreement required	Yes

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