# CHO-CD36

305979

## GENERAL INFORMATION

### Description

Disclaimer: The prices displayed for cell lines are exclusively for academic/not-for-profit customers. For commercial entitites the price is approximately €6,250.
If you represent a commercial entity or are unsure which category applies, please
contact us.
CHO-CD36 cells are recombinant Chinese hamster ovary (CHO) cells engineered to stably express human CD36, a multifunctional class B scavenger receptor also known as platelet glycoprotein IV (GPIV) or fatty acid translocase (FAT). CD36 is broadly involved in lipid uptake, fatty acid metabolism, angiogenesis, inflammation, innate immunity, and cell adhesion. The receptor interacts with a wide range of ligands, including oxidized low-density lipoproteins (oxLDL), long-chain fatty acids, thrombospondin-1, phospholipids, and apoptotic cells. Dysregulated CD36 expression has been implicated in metabolic disorders, atherosclerosis, chronic inflammation, and tumor progression, making recombinant CD36-expressing cell models valuable tools for mechanistic and therapeutic research.
CHO-CD36 cells are widely used for studying receptor-ligand interactions, lipid transport mechanisms, and therapeutic targeting of CD36-associated pathways. These cells support quantitative analysis of ligand binding, receptor internalization, fatty acid uptake, and downstream signaling events linked to oxidative stress, immune modulation, and metabolic adaptation. In oncology research, CHO-CD36 models are useful for investigating the role of CD36 in metastasis, tumor lipid metabolism, and resistance to metabolic stress. The cells are also applied in development and characterization of monoclonal antibodies, small-molecule inhibitors, lipid-targeting therapeutics, and imaging agents directed against CD36. Flow cytometry assays, uptake assays, and high-throughput screening platforms commonly utilize CHO-CD36 cells because of their stable and controlled recombinant receptor expression.

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| **Tissue** | Ovary |
| **Disease** | Chinese hamster ovary, non-neoplastic; genetically engineered for CD36 surface expression |

### Applications

Antibody screening; CD36-targeted therapy development; lipid metabolism research; scavenger receptor biology; flow cytometry

## CLASSIFICATION

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| **Organism** | Chinese hamster |
| **Sex** | Female |
| **Growth properties** | Adherent |
| **GMO status** | GMO-S1 |

### GMO status details

GMO-S1: This CHO cell line contains a CD36 expression cassette supporting receptor-function analyses. This classification applies only within Germany and may differ elsewhere.

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| **Biosafety level** | 1 |

## CHARACTERISTICS

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| **Morphology** | Epithelial-like |
| **Cell type** | Epithelial cell of ovary |
| **Doubling time** | approx. 14-16 hours |

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## IDENTIFIERS

**Citation**

CHO-CD36 (Cytion catalog number 305979)

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| **NCBI TaxID** | 10029 |
| **Cellosaurus** | CVCL_8848 |

## EXPRESSION AND SUSCEPTIBILITY

**Receptors expressed**

CD36

## HANDLING AND CULTURE

**Culture medium**

For adherent cultures: DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO3 (Cytion article number 820400a)
For suspension cultures: CHO Growth Medium A (from InSCREENeX; InSCREENeX catalog number INS-ME-1039)

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| **Incubation atmosphere** | 37°C, 5% $CO_2$, humidified atmosphere. |

**Supplements**

For adherent cultures: Supplement the medium with 5% FBS. Add Geneticin (G418-Sulfat) to achieve a final concentration of 0.5 mg/mL.

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| **Dissociation reagent** | For adherent cultures: Trypsin-EDTA |

**Subculturing**

For routine adherent cell culture: Aspirate the old culture medium from the adherent cells, and wash them with PBS to remove any remaining medium. After aspirating the PBS, add the appropriate volume of Trypsin/EDTA solution based on the culture vessel size (e.g., 1 ml for a T25 flask, 3 ml for a T75 flask) and incubate at room temperature or 37°C for 5-10 minutes, or until the cells detach. Monitor detachment under a microscope, and gently tap the vessel if necessary to release the cells. Once detached, add complete medium to inactivate the Trypsin/EDTA, gently resuspend the cells, and transfer an aliquot of the cell suspension into a new culture vessel containing fresh medium. Place the vessel in an incubator set to 37°C with 5% $CO_2$, and change the medium every 2-3 days.

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| **Split ratio** | 1 to 5 |
| **Seeding density** | 2 to 5 x $10^4$ cells/$cm^2$ |
| **Fluid renewal** | 2 to 3 times per week |
| **Post-thaw recovery** | After thawing, split the cells at a ratio of 1:2 to 1:3 in T25 flasks and allow the cells to recover from the freezing process and to adhere (for adherent cultures) for at least 24 hours. |

**Thawing and culturing**

Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

## QUALITY CONTROL AND CRYOPRESERVATION

### Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.
To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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| **Freeze medium** | As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress. |

### Storage conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

### Shipping conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

## DONOR

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| **Donor age** | Adult |

## LEGAL AND SAFETY

### Safety precautions

When planning to store a cryovial in liquid nitrogen for future thawing, it is mandatory to adhere to stringent safety measures. Appropriate protective gloves and clothing are essential, and the use of a face mask or safety goggles is required during the transfer of frozen samples to or from the liquid nitrogen tank. This is to mitigate the risk of injury from potential cryovial explosions upon removal, which can result in the projection of sharp fragments.

### Warranty

At Cytion, we stand behind the quality and viability of our products.
We guarantee that our cells will be viable at the start of culture for
60 days after shipment from our facilities.
We assure that all materials conform to the specifications described in the
Certificate of Analysis and/or Product Sheet.
For full details, please refer to our
Warranty Statement.

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| **Third-party agreements apply** | Yes |